

SCANNING ELECTRON MICROSCOPIC INVESTIGATIONS ON THE  
LEAF MICRO-MORPHOLOGICAL AND MESOPHYLL PARAMETERS IN  
ASSOCIATION WITH PHOTOSYNTHETIC ACTIVITY OF THREE  
MULBERRY (*MORUS* SPP.) GENOTYPES

Vineet K.<sup>1,\*</sup>, Kodandaramaiah J.<sup>2</sup>, Jhansi Lakshmi K.<sup>1</sup>, Mala V. Rajan<sup>2</sup>  
and Kamble C.K.

<sup>1</sup>Electron Microscopy Division, <sup>2</sup>Mulberry Physiology Laboratory  
Central Sericultural Research and Training Institute  
Srirmapura, Mysore – 570008, India.

\*Corresponding author email ID: [vinkumar2006@rediffmail.com](mailto:vinkumar2006@rediffmail.com)

ABSTRACT

A scanning electron microscopic study on the micro-morphological and mesophyll parameters of three mulberry genotypes having different yield potentials viz. Punjab local, S-30 and China Peking was under taken to find out their association with photosynthetic rate. The results of the present study revealed that the leaf thickness of Punjab local was found maximum ( $127.42 \pm 0.582 \mu\text{m}$ ) and conversely minimum in S-30 genotype ( $119.17 \pm 0.752 \mu\text{m}$ ). Statistically significant differences were found among three genotypes for leaf thickness character against all the other parameters like thickness of palisade and spongy tissues, percentage of palisade parenchyma in mesophyll and the rate of photosynthesis. With regard to the total density of idioblasts per  $\text{mm}^2$  leaf surface area, it was observed that sufficient variability exist among the genotypes. The experimental data also indicated the highest photosynthetic rate in S-30 genotype ( $21.56 \pm 0.357 \mu \text{mol m}^{-2} \text{s}^{-1}$ ) with contrastingly low in China Peking ( $11.06 \pm 0.529 \mu \text{mol m}^{-2} \text{s}^{-1}$ ) significantly associating with the respective micro-morphological and mesophyll characteristics. Hence, these structural traits which are found correlated with higher physiological efficiency can be conveniently utilized in the screening and evaluation of mulberry genotypes for further improvement in leaf yield and biomass production by the plant breeding programmes.

**Keywords:** micro-morphological parameters, mulberry genotypes, photosynthetic rate, PAR, SEM

INTRODUCTION

Mulberry (*Morus* spp.), the only food source of silkworm, *Bombyx mori* Linn., is of great economic importance to the sericulture industry. The quality and quantity of the leaf produced has a direct bearing on the cocoon production and in turn, the economy of the rural farmers. The different factors responsible for a successful cocoon crop are: mulberry leaf (38.2%), climatic conditions (37.0%), rearing techniques (9.3%), silkworm races (4.2%), silkworm eggs (3.1%) and other factors

(8.2%). Therefore, the economics and profitability of sericulture depends primarily on quality of the mulberry leaves produced (Singhal et al. 1999). Genetic and environmental factors influence the growth and quality of mulberry leaves. The effect of water stress on photosynthetic activity in various crops has been well established (Ackerson et al. 1977; Hutmacher & Krieg 1983; Ephrath et al. 1990; Srinivasa Rao & Bhatt 1990; Leidi et al. 1993). Chakrabarti et al. (1996) described the relationship between gas exchange traits and genetic variability in mulberry under irrigated and rainfed conditions. Recently, Babu et al. (2006) have reported the adaptive anatomical characteristics of mulberry leaves genotypes for shade tolerance.

During the evaluation and selection of various mulberry genotypes for the breeding programmes, different morpho-physiological parameters are considered for developing high yielding mulberry varieties. However, it is practically difficult to consider all genotypes for selection and, therefore, they have to be short listed from a large number of progenies based on their easily measurable and useful characters. The literature clearly points out that the prime physiological parameter namely photosynthesis, which is the basis for biological yield, is correlated to stomata frequency for gas exchange and, thus has direct effect on the crop production.

An attempt is, therefore, made to investigate leaf micro-morphology and anatomy by scanning electron microscopy of three mulberry genotypes which may help in identifying certain crucial characters before taking up the mulberry improvement programmes. The survey of literature reveals that such type of studies have not been carried out so far. The ultimate objective of the study is to identify micro-morphological/anatomical and physiological traits which may help in the selection process during early stage of evaluation for short-listing the genotypes during breeding programmes as mulberry is unique with large genetic variability and long gestation period.

## **MATERIALS AND METHODS**

### **Cultivation of mulberry**

For the present study, three mulberry genotypes namely China Peking, Punjab Local and S-30 were selected based on low, medium and high yield potential from the germplasm resource bank of Central Sericultural Research and Training Institute, Mysore, India. These mulberry genotypes were established in random block design (RBD) in plots of size 5.4 m by 3.6 m under irrigated conditions with standard fertilizers (300:120:120 kg NPK/ha/y) and manure application (20 mt compost/ha/y) (Dandin et al. 2000). The spacing between rows and plants was 120 cm x 120 cm. Five replications were maintained per genotype in four different plots. The garden soil used was red loamy with pH of 7.25 to 7.65 and electrical conductivity of 0.11 to 0.13 m mhos/cm. The fully matured mulberry leaves of all the three genotypes

were collected 70 days after bottom pruning was conducted as per standard sericultural practices and used for SEM study.

### **Procedure for scanning electron microscopy**

To study the leaf thickness, palisade and spongy parenchyma and stomata frequency under scanning electron microscope (SEM), the samples of 3 mm x 3 mm, and 3 mm x 1 mm sizes were fixed for 4 hours in glutaraldehyde prepared in 0.2 M sodium cacodylate buffer (pH 7.2). The fixed samples were washed thrice in sodium cacodylate buffer and then dehydrated in an alcohol-acetone series. The dehydrated samples were dried in a critical point drier (EMS - 850) using CO<sub>2</sub> as a transition fluid. The dried samples were mounted on to copper stubs using double side stick cellophane tape. To observe cross sectional area, the 3 mm x 1 mm sized leaf tissues were vertically mounted exposing their cut surfaces. The dried samples were gold coated (20 nm thickness) in a sputter coater (EMS - 550). The coated samples were observed under a transmission electron microscope (JEOL 100 CX II, Tokyo Pvt. Ltd. Japan) attached with scanning device (ASID 4 D) at 20 kV, and photographs were taken at different magnifications. Five samples from each genotype were examined for confirmation of the results. The thickness of leaf and mesophyll tissues was recorded from SEM micrographs and statistically analyzed.

### **Photosynthetic rate**

Photosynthetic rate was measured in all three genotypes *viz.*, Punjab Local, S-30 and China Peking between 10 to 12 hours under natural sun light using a portable photosynthetic system (Model-LI 6200, LI-COR Co. Ltd. Lincoln, NE, USA). The photosynthetically active radiation (PAR) was around 1500  $\mu\text{mol m}^{-2} \text{s}^{-1}$  during the gas exchange measurements. The fully matured and indexed 12<sup>th</sup> leaf from the top of the shoot was selected, and the gas exchange data was recorded at the time of harvest (70 days).

### **Statistical analysis**

The data were subjected to statistical analysis employing analysis of variance (ANOVA) to ascertain the significance of various characters *viz.* leaf thickness, thickness of palisade and spongy parenchyma, percentage of palisade parenchyma in mesophyll tissue, photosynthetic rate, number of stomata (per  $\text{mm}^2$ ), number of trichomes (per  $\text{mm}^2$ ), and number of idioblasts (per  $\text{mm}^2$ ) among the three mulberry genotypes.

## **RESULTS AND DISCUSSION**

### **Mesophyll characteristics**

Results of the scanning electron microphotographs of transverse section of mulberry leaves of three mulberry genotypes *viz.* Punjab local, S-30 and China Peking

showed variation in leaf thickness, palisade tissue, and spongy parenchyma (Figure 1-6). The leaf thickness of mulberry genotype Punjab local recorded at  $127.42 \pm 0.582 \mu\text{m}$  (Figure 1-2) whereas it was  $119.17 \pm 0.752 \mu\text{m}$  and  $119.61 \pm 0.623 \mu\text{m}$  for genotypes of S-30 (Figure 3-4) and China Peking (Figure 5-6), respectively (Table 1).

Table 1. Variation in mesophyll characters and the rate of photosynthesis of three different mulberry genotypes.

Genotype	Leaf thickness	Thickness of palisade parenchyma ( $\mu\text{m}$ )	Thickness of spongy parenchyma ( $\mu\text{m}$ )	Percentage of palisade parenchyma in mesophyll ( $\mu\text{m}$ )	Photosynthetic rate ( $\mu\text{mol m}^{-2}\text{s}^{-1}$ )
Punjab Local	$127.42 \pm 0.582$	$62.46 \pm 0.941$	$50.77 \pm 0.753$	$50.41 \pm 0.660$	$17.09 \pm 0.088$
S-30	$119.17 \pm 0.752$	$46.72 \pm 0.371$	$36.54 \pm 0.649$	$56.05 \pm 0.396$	$21.56 \pm 0.357$
China Peking	$119.61 \pm 0.623$	$43.22 \pm 0.444$	$59.69 \pm 0.703$	$41.94 \pm 0.152$	$11.06 \pm 0.529$
Standard Error	0.747	0.635	0.680	0.468	0.332
F Test	***	***	***	***	***
CD at 5% level	2.435	2.072	2.018	1.526	1.083

$\pm$  = Standard Error

\*\*\* Significant at 0.01 level

Babu et al. (2006) while investigating the three different mulberry genotypes under full sunlight and shade conditions have found that the thickness of V1 genotype in sunlight was  $104.2 \mu\text{m}$  and  $45.8 \mu\text{m}$  in the shade, whereas it was  $93.8$  and  $114.6 \mu\text{m}$  in sunlight and  $72.9$  and  $79.2 \mu\text{m}$  in shade for K2 and K2 x Kosen genotypes. Further, it was also reported that the leaves of sun-exposed plants tend to be thicker than those of the shade-grown ones (Igboanugo 1993; Mendes et al. 2001), and the light demanding species have the maximum leaf anatomical plasticity (Ashton & Berlyn 1994). When the SEM data of present study for leaf

thickness of three mulberry genotypes were statistically analyzed, there was significant difference among Punjab local, S-30 and China Peking. However, leaf thickness character did not differ between the genotypes of S-30 and China Peking (Table 1). Nonetheless, data for other traits such as thickness of palisade and spongy tissues, percentage of palisade parenchyma in mesophyll, and the rate of photosynthesis, in all the genotypes were found highly significant (Table 1).

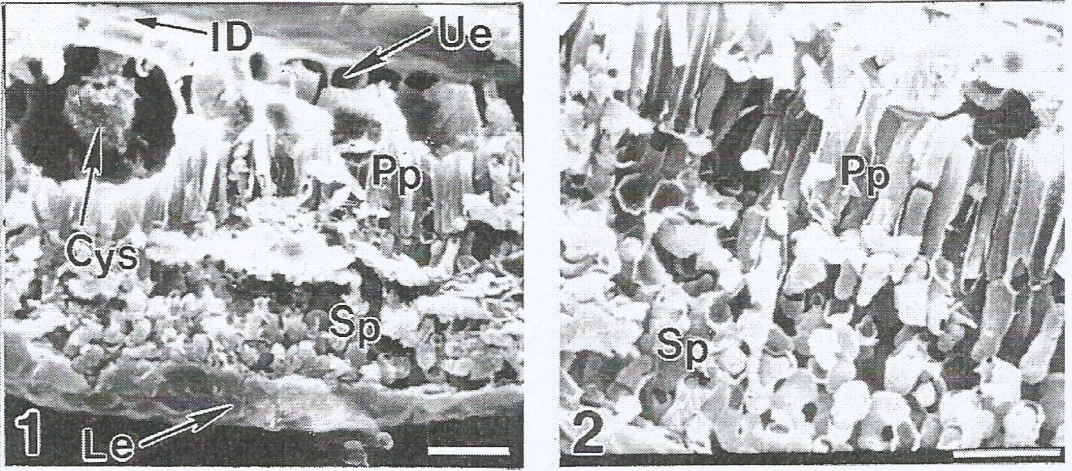


Figure 1-2. Microphotographs of the transverse section of mulberry leaf of Punjab local genotype revealing the upper epidermis (Ue), palisade parenchyma (Pp), spongy parenchyma (Sp) and lower epidermis (Le). Note idioblast (ID) opens in a cystolyth (Cys) just below the upper epidermis. Scale bar = 12  $\mu$ m for Figure 1, and 10  $\mu$ m for Figure 2.

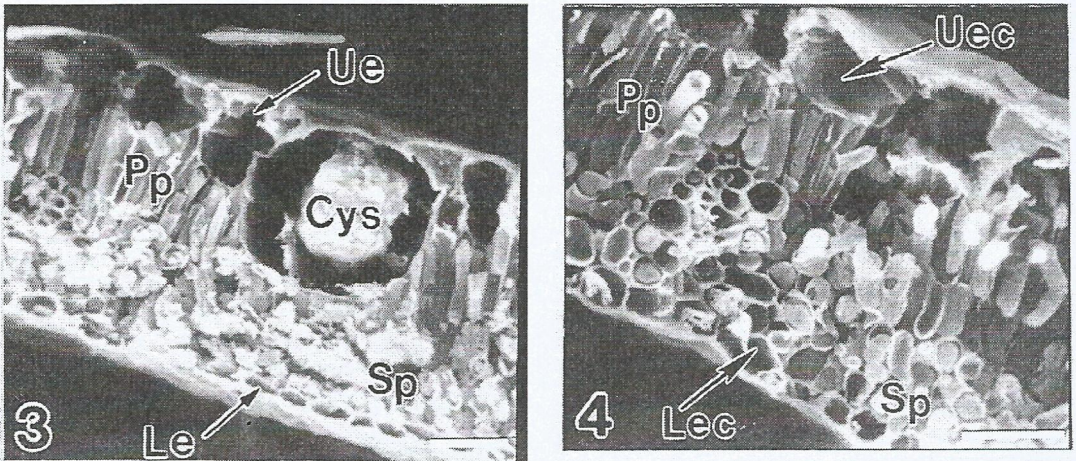


Figure 3-4. Transverse section of mulberry leaf of S-30 genotype revealing the upper epidermis (Ue), palisade parenchyma (Pp), spongy parenchyma (Sp) and lower epidermis (Le). Scale bar = 12  $\mu$ m for Figure 3, and 10  $\mu$ m for Figure 4.

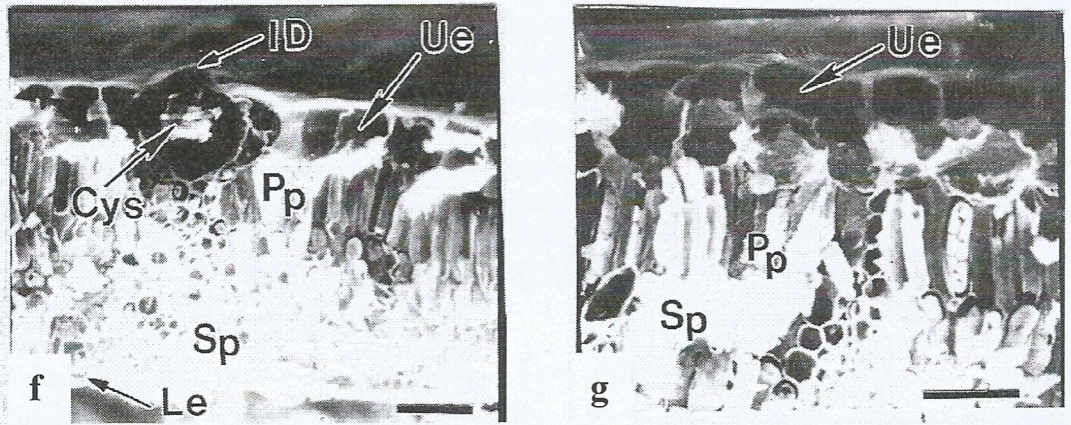


Figure. 5-6. Transverse section of mulberry leaf of China Peking genotype revealing the upper epidermis (Ue), palisade parenchyma (Pp), spongy parenchyma (Sp) and lower epidermis (Le). Note idioblast (ID) opens in a cystolyth (Cys) just below the upper epidermis. Scale bar = 12  $\mu\text{m}$  for Figure 5, and 10  $\mu\text{m}$  for Figure 6.

It is well documented that palisade parenchyma is an important character of the leaves of any plant as it reflects better ability of the plant genotypes to capture solar energy and accounts for photosynthetic efficiency. The thickness of palisade parenchyma in Punjab local genotype (Figure 1-2) was found to be  $62.46 \pm 0.941 \mu\text{m}$  whereas it was  $46.72 \pm 0.371 \mu\text{m}$  and  $43.22 \pm 0.444 \mu\text{m}$  for S-30 (Figure 3-4) and China Peking (Figure 5-6), respectively. There was a significant difference for the character of the thickness of palisade parenchyma among three genotypes studied (Table 1). Further, the values of leaf thickness of three genotypes were highly significant against all the other characters of mesophyll tissue and photosynthetic rate (Table 1). Babu et al. (2006) have earlier studied the percentage of palisade tissue of three mulberry varieties and the data of the leaves grown in sunlight and shade conditions were documented as 48.0% and 40.8% (V1); 37.7% and 28.5% (K2) and 45.4% and 50.0% (K2 x Kosen), respectively.

The healthy leaves of the three genotypes had an almost similar internal structure typical of a dorso-ventral dicot leaf with prominent distinguishable palisade and spongy parenchyma in the mesophyll. However, the thickness of spongy parenchyma distinctly differed among the three genotypes studied. The mean values of the spongy parenchyma of three genotypes were shown in Table 1. It was found maximum in China Peking genotype ( $59.69 \pm 0.703 \mu\text{m}$ ) whereas it was least observed in S-30 genotype ( $36.54 \pm 0.649 \mu\text{m}$ ). The statistical analysis showed a significant difference among the three genotypes for the thickness of spongy parenchyma (Table 1).

Since the mesophyll structure is associated with photosynthetic performance of leaves via the regulation of internal light and  $\text{CO}_2$  profiles (Babu et

al. 2006), the percentage of palisade parenchyma is a crucial trait in the mesophyll tissue of mulberry genotypes. During the present study the percentage of palisade parenchyma was found highest i.e.  $56.05 \pm 0.396 \mu\text{m}$  in S-30 genotype while the values were recorded minimum for China Peking ( $41.94 \pm 0.152 \mu\text{m}$ ). Further, the data revealed a highly significant difference between the three genotypes for the percentage of palisade tissue in mesophyll (Table 1).

### **Rate of photosynthesis**

Leaf yield of any plant generally depend on the photosynthetic rate, which plays a vital role in biomass productivity. Therefore, it is necessary to consider this prime physiological parameter for mulberry improvement programme. Screening of progeny and short listing by evaluation is the primary step in identification of genotypes in the initial stage for inclusion in primary yield trait in any breeding programme. A rapid screening method is essential to reduce the time and labour required in identification of suitable genotypes for a particular condition (Fotadar et al. 2006). The experimental data revealed that the highest photosynthetic rate recorded was  $21.56 \pm 0.357 \mu\text{mol m}^{-2} \text{s}^{-1}$  in S-30 genotype whereas Punjab local stood at second place with the values of  $17.09 \pm 0.088 \mu\text{mol m}^{-2} \text{s}^{-1}$ . Genotype China Peking was noticed with lowest photosynthetic rate i.e.  $11.06 \pm 0.529 \mu\text{mol m}^{-2} \text{s}^{-1}$ . The CD values for the photosynthetic rates measured showed a significant difference among the genotypes (Table 1).

### **Micro-morphological characteristics of abaxial leaf surface**

A scanning electron microscopic study was carried out to find out the micro-morphological characters like density of stomata, trichomes and idioblasts per unit surface area of three mulberry genotypes and the data were subjected to statistical analysis (ANOVA) and presented in Table 2. The average number of stomata was found  $895 \pm 6.804$ ,  $887 \pm 6.253$   $425 \pm 3.536$  per  $\text{mm}^2$  in Punjab local, S-30 and China Peking respectively. Babu et al. (2006) reported a significant reduction in the number of stomata on the leaves of mulberry plants grown under shade condition. Further, a comparatively less stomata frequency in the leaves of shade plants than the sun exposed ones was fairly documented in many plants species (Rocas et al. 1997; Mendes et al. 2001). Since the leaf surface structure is associated with the photosynthetic performance via regulation of internal light and  $\text{CO}_2$  profiles, the genotype China Peking seems not to be much useful for mulberry productivity improvement programmes as it has minimum number of stomata per  $\text{mm}^2$ . On the other hand, Punjab local and S-30 genotypes can be useful for breeding purposes as both the genotypes have more stomata frequency. Though the data is not showing significant difference between the genotypes of Punjab local and S-30, yet the genotype China Peking was found significant when compared to others.

Table 2. Variation in number of stomata, trichomes and idioblasts in three mulberry genotypes.

Genotypes	Number of stomata (Per mm <sup>2</sup> )	Number of Trichomes (Per mm <sup>2</sup> )	Number of idioblasts (Per mm <sup>2</sup> )
Punjab Local	895±6.804	112.5±5.853	67.0±3.364
S-30	887±6.253	140.4±3.855	29.0±2.702
China Peking	425±3.536	168.6±5.212	34.0±2.270
Standard Error	6.379	5.474	1.522
F Test	***	***	***
CD at 5% level	20.805	17.851	4.964

± = Standard Error

\*\*\* Significant at 0.01 level

Plant species exhibit wide variation in types and densities of trichomes within the families. Trichomes are the epidermal appendages that consist of one or more cells derived from a single proto-dermal cell (Kesavacharyulu et al. 2004). In many plants, trichomes have been used to classify genera and species (Metcalf & Chalk 1979; Mehta et al. 1979) including mulberry (Katsumata 1971; Fujita & Uchikawa 1986). The role of trichomes in plant defense against herbivore is known since long (Callahan 1957; Beck 1965; Levin 1973; Norris & Kogan 1980; Jermy 1984). The foliar trichomes when present in high density are reported to cause physical hindrance and discourage phytophagous insects, and affects acceptability of foliage (Singh et al. 1971; Levin 1973).

Trichomes play a major role in acceptability as feed by insects (Kesavacharyulu et al. 2004) and in some cases were reported to synthesize, metabolize, accumulate, and secrete a variety of substances (Norris & Kogan 1980; William et al. 1984; Purushothaman & Vasanth 1988; Pedro et al. 1991; Werker 1993). The secretions from glandular trichomes probably play a role to attract the pollinators (Ascensao et al. 1995). In the present study, the glandular trichomes were present on the abaxial surface of three mulberry genotypes were recorded as 112.5±5.853 per mm<sup>2</sup> in Punjab local (Figure 7-8), 140.4±3.855 per mm<sup>2</sup> in S-30 (Figure 9-10), and 168.6 5.212 per mm<sup>2</sup> in China Peking (Figure 11-12).



Significant differences were observed among these three genotypes (Table 2; Figures 7-12).

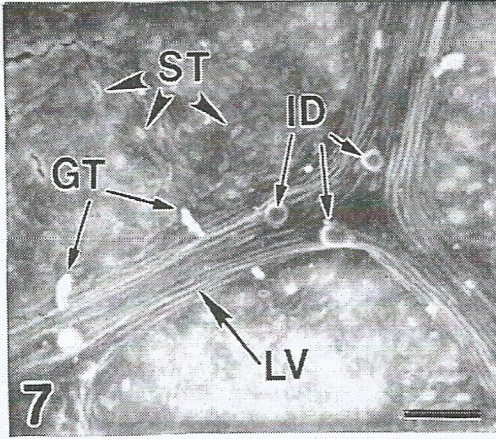


Figure 7. Abaxial leaf surface of mulberry genotype of Punjab local showing the leaf vein (LV), glandular trichomes (GT), idioblast (ID) and stomata (ST). Scale bar = 30  $\mu$ m.

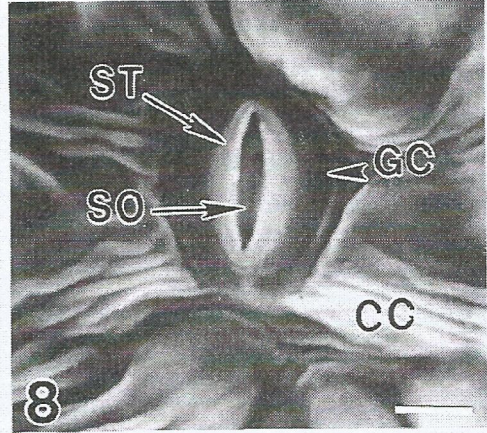


Figure 8. A magnified view of stomata of Punjab local reveals the stomata opening (SO), guard cell (GC) and companion cell (CC). Scale bar = 2  $\mu$ m.

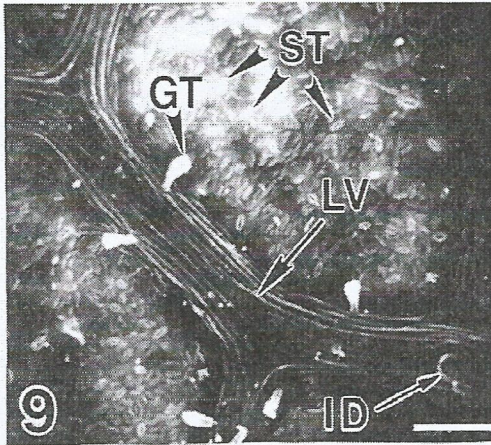


Figure 9. Abaxial leaf surface of mulberry genotype of S-30 showing the leaf vein (LV), glandular trichomes (GT), idioblast (ID) and stomata (ST). Scale bar = 30  $\mu$ m.

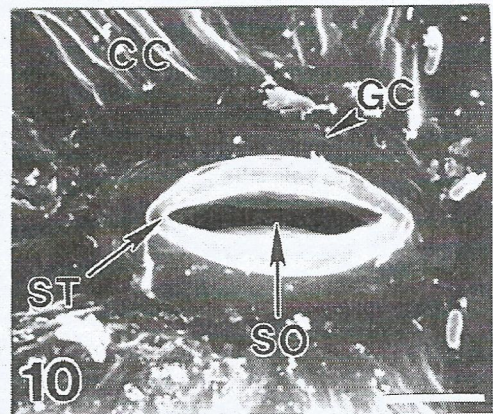


Figure 10. Microphotograph reveals the stomata (ST), stomata opening (SO), guard cell (GC) and companion cell (CC). Scale bar = 2  $\mu$ m.

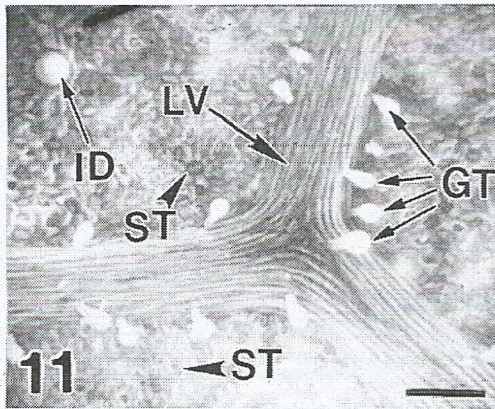


Figure 11. Abaxial leaf surface of mulberry genotype of China Peking showing the leaf vein (LV), glandular trichomes (GT), idioblast (ID) and stomata (ST). Scale bar = 30  $\mu\text{m}$ .

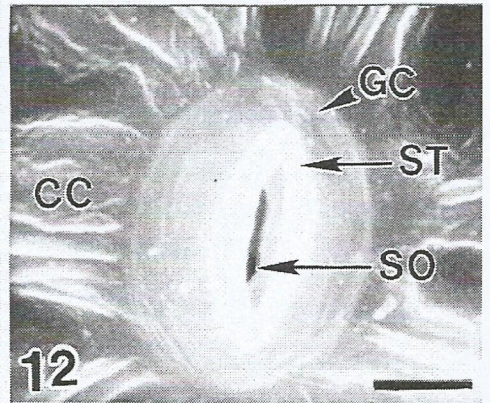


Figure 12. A magnified view of stomata of the genotype of China Peking reveals the stomata opening (SO), guard cell (GC) and companion cell (CC). Scale bar = 2  $\mu\text{m}$ .

While considering the total density of idioblasts per  $\text{mm}^2$  leaf surface area, it was observed that sufficient variability exists among these genotypes. The density of idioblasts in genotype Punjab local recorded maximum values ( $67.0 \pm 3.364$ ) and lowest in S-30 ( $29.0 \pm 0.702$ ), whereas the number was medium in China Peking ( $34.0 \pm 2.270$ ; Figures 7-12). There was a significant difference in number of idioblasts among three mulberry genotypes studied.

## CONCLUSION

Overall, the results of the present SEM investigation revealed that the differences in micro-morphological and mesophyll characteristics are directly associated with variability in light interception and photosynthetic efficiency of mulberry genotypes. These micro-morphological and photosynthetic traits can prove their potential in the selection and screening of improved mulberry varieties in a relatively shorter span of time.

## REFERENCES

- Ackerson RC, Krieg DR, Haring CL, Chang N. 1977. Effects of plant water status on stomatal activity, photosynthesis and nitrate reductase activity of field grown cotton. *Crop Science* 17: 81-84.

- Ascensao L, Marques N, Pais MS. 1995. Glandular trichomes on vegetative and reproductive organs of *Leonotis leonurus* (Lamiaceae). *Annals of Botany* **75**: 619-626.
- Ashton PMS, Berlyn GP. 1994. A comparison of leaf physiology and anatomy of *Quercus* (section *Erythrobalanum* – Fagaceae) species in different light environments. *American Journal of Botany* **81**: 589-597.
- Babu AM, Guha A, Kumar JS. 2006. Adaptive anatomical characteristics of leaves of three mulberry genotypes for shade tolerance. *Indian Journal of Sericulture* **45**(1): 76-80.
- Beck SD. 1965. Resistance of plants to insects. *Annual Review of Entomology* **10**: 207-232.
- Callahan PS. 1957. Oviposition response of the corn earworm to differences in surface texture. *Journal of the Kansas Entomological Society* **30**: 59-63.
- Chakrabarti S, Singhal BK, Thippeswamy T, Rekha M. 1996. Studies on the relationship between gas exchange traits and genetic variability in mulberry (*Morus alba* L.) under irrigated and rainfed conditions. *Indian Journal of Sericulture* **35**(1): 54-56.
- Dandin SB, Jayaswal J, Gridhar K. 2000. *Hand Book of Sericulture Technologies*. Central Silk Board. Bangalore, India. pp. 39.
- Ephrath JE, Marani A, Bravado BA. 1990. Effects of moisture stress on stomatal resistance and photosynthetic rate in cotton (*Gossypium hirsutum* ) I. Controlled levels of stress. *Field Crops Research* **23**: 117-131.
- Fotadar RK, Sengupta D, Khan MA. 2006. Nursery evaluation technique for preliminary selection of genotypes under sub-tropical conditions of Jammu. *Indian Journal of Sericulture* **45**(1): 81-84.
- Fujita H, Uchikawa. 1986. Electron microscopical study of mulberry with special reference to the identification of cultivars: In Kitaura K, Horie M, Kozaki (eds) *Development Of New Technology For Identification And Classification Of Tree Crops And Ornamentals* . I. Fruit Tree Research Station, Ministry of Agriculture, Forestry and Fisheries, Japan. pp 25-29.
- Hutmacher RB, Krieger DR. 1983. Photosynthetic rate control in cotton. Stomatal and non-stomatal factors. *Plant Physiology* **73**: 658-661.
- Igboanugo ABT. 1993. Analysis of leaf development under three irradiance levels in a shade tolerant and shade non-tolerant trees species. *Nigerian Journal of Botany* **6**: 91-102.
- Jermey T. 1984. Evolution of insect/host plant relationships. *The American Naturalist* **124**: 609-630.
- Katsumata F. 1971. Shape of idioblasts in mulberry leaves with special reference to the classification of mulberry trees. *Journal of Sericulture Science of Japan* **40**: 313-322.

- Kesavacharyulu K, Kumar V, Sarkar A. 2004. Scanning electron microscopic studies on leaf surface trichomes in mulberry and its influence on rearing performance of silkworm *Bombyx mori* L. *International Journal of Industrial Entomology* **8**(1): 33-41.
- Leidi EO, Lopez JM, Lopez M, Gutierrez JC. 1993. Searching for tolerance to water stress in cotton genotypes: photosynthesis, stomatal conductance and transpiration. *Photosynthetica* **28**: 383-390.
- Levin DA. 1973. The role of trichomes in plant defense. *The Quarterly Review of Biology* **48**: 3-15.
- Mehta IJ, Dhillon PS, Hanson GP. 1979. Trichome morphology as an indicator of high rubber bearing guayule (*Parthenium argentatum* Gray) Plants in native populations. *American Journal of Botany*. **66**: 769-804.
- Mendes MM, Gazarini, LC, Rodrigues ML. 2001. Acclimation of *Myrtus communis* to contrasting Mediterranean light environments: Effects on structure and chemical composition of foliage and plant water relations. *Environmental and Experimental Botany* **45**: 165-178.
- Metcalf CR, Chalk L. 1979. *Anatomy of the Dicotyledons* . Second edition Vol 1. Systematic anatomy of leaf and stem, with a brief history of the subject. Clarendon Press, Oxford.
- Norris DM, Kogan M. 1980. Biochemical and morphological bases of resistance. In Maxwell FG, Jennings PR (eds) *Breeding Plants Resistant to Insects* . John Wiley & Sons, New York. pp. 23-62.
- Pedro LG, Barroso JG, Marques NT, Ascensao L, Pais MS, Scheffer JJ. 1991. Composition of essential oil from sepals of *Leonotis leonurus* R. Br. *Journal of Essential Oil Research* **3**: 451-453.
- Purushothaman KK, Vasanth S. 1988. Chemical studies on *Leonotis*. *A Review of Indian Drugs* **25**: 484-491.
- Rocas G, Barros CF, Scarano FR. 1997. Leaf anatomy plasticity of *Alchornea triplinervia* (*Euphorbiaceae*) under distinct light regimes in a Brazilian montane Atlantic rain forest. *Trees Structure and Function* **11**: 469-473.
- Singh BB, Hardley HH, Bernard RL. 1971. Morphology of pubescence in Soybeans and its relationship to plant vigour. *Crop Science* **11**: 13-16.
- Singhal BK, Rajan MV, Sarkar A, Datta RK. 1999. Nutritional disorders of mulberry (*Morus* Spp.) III. Leaf nutrient guide for secondary nutrients. *Sericologia* **39**(4): 599-609.
- Srinivasa Rao NK, Bhatt RM. 1990. Response of photosynthesis to water stress in two egg plant (*Solanum melongena* L.) cultivars. *Photosynthetica* **24**: 506-513.

- Werker E. 1993. Function of essential oil secreting glandular hairs in aromatic plants of Lamiaceae - A review. *Flavor and Fragrance Journal* **8**: 249-255.
- William WT, Healey PI (1984) Cellular basis of trichome secretion; In Rodriguez, B, Healey PI, Mehata I (eds) *Biology and Chemistry of Plant Trichomes* , Plenum Press, New York. pp 95-111.